

## A Comparative Study of Nutritive Value In Fresh And Salt Dried Fish

### <sup>1</sup>Dr.Remya V K, <sup>2</sup>Vinitha M S, <sup>3</sup>Jijisha T S

<sup>2</sup>Assistant Professor in Zoology1 Post Graduate Department of Zoology, Sree Narayana College,Nattika Thrissur, Kerala,India.

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**ABSTRACT:** The present study investigates the comparion of nutritive values fresh and salt dried fishes. The two fish species selected are Rastrelliger kanagurta and Sardinella longiceps. The protein carbohydrate and lipid content were by methods Lowry analyzed method, Phenolsulphric acid method and Bligh&Dyer method respectively. The resultsshows that fresh fish has higher nutritional value when compared with salt dried fish. Rastrelliger kanagurta and Sardinella longiceps, this is because of salt causes peroxidation reactions.

Key words: Fish, Nutrition, Salt dry

#### I. INTRODUCTION

Fish preservation is a very important aspect of the capture and culture fisheries. If it not done properly after the capture, the very purpose of their raising is lost.normally the fish farms and other fish capturing sites are located far from the market place and transportation of the fish presents problems like the speed at which the fish decomposes and the uncertainties of their sale in the market. When at times the fishes are caught in numbers greater than the amount of consumption, their preservation becomes a necessity for their future use.drying as a means of preserving fish, has been practiced perhaps longer than any other food preservation technique.it involves dehydration so that the bacterial decomposition of enzymatic autolysis does not occur.sun drying is the most prevalent preservation method in India.

Fish is an important source of a healthy diet and is considered as the biggest source of protein. By composition fish, contain fat, free amino acids, and water.fish is a good dietary source for fat-soluble vitamins. The macro elements present in fish are calcium, phosphorus, magnesium, sodium, potassium, chlorine.from a nutritional point of view, protein is the most important constituent of fish, which determine its wholesomeness and quality. Major lipids are triacylglycerols and phospholipids.fish accumulates the highly unsaturated fatty acids, notably n-3 pufa's. Fish contain a trace amount of carbohydrate, which is insignificant from the nutritional point of view.but they are the important factor affecting the quality of fish during processing.

#### THE OBJECTIVE OF THE STUDY

Comparitive of nutritive value of fresh and salt dried fish.

#### **II. MATERIALS AND METHODS**

For analysis two species of fishes were selected. These two species are daily used food fishes. Both fishes are collected from the local fish market of Nattikka.flesh of fish separated and then subjected to carbohydrate, protein, and lipid analysis.

Two species of fishes were selected. Rastrelliger kanagurta and Sardinella longiceps

Protein estimation by Lowry Method

The Lowry protein assay is a biochemical assay for determining the total level of protein in a solution. Principle: alkaline cuso4 catalyzes the oxidation of aromatic amino acids with subsequent reduction of sodium-potassium molybdate tungstate of Folin's reagent giving a purple color complex the intensity of the color is directly proposition to the concentration of the aromatic amino acid in the given sample solution.

Reagents required:

1. Stock solution: bovine serum albumin of 100mg is weighed accurately and dissolved in 100ml of distilled water in a standard flask (concentration 1  $\mu$ g/ml).

2. Working standard: the stock solution of 10 ml is distilled to 100ml with distilled water in a standard flask (concentration 100 mg/ml).

3. Folin's phenol reagent: Folin's phenol reagent is mixed with distilled water in the ratio 1:2.



4. Alkaline copper reagent: solution a: 2% sodium carbonate in 0.1 n sodium hydroxide. Solution b: 0.5% copper sulphate in 1% sodium potassium tartrate. Solution a, b, c is mixed in the proportion of 50:1:0.

5. Unknown preparation: the unknown protein is made up to 100 ml with distilled water.

Procedure: working standard of 0.2 -1ml is pipette out into a clean test tube and labeled as s1-s5. A test solution of 0.2ml is taken into the test tube and labeled as t1. The volume is made up to 1ml of distilled water. Distill water of 1ml serves as blank. To all the test tubes 4.5ml of alkaline cuso4 reagent is added and incubated at room temperature for 10 minutes. All the test tube 0.5ml of Folin's phenol reagent is added. The contents are mixed well and the blue color developed is read at 640 rpm after 15 minutes. From the standard graph, the amount of protein in the given unknown solution is calculated.

Lipid estimation- Bligh/Dyer extraction of total lipids:

#### Reagents

1.Chloroform-methanol reagent

2.0.05n potassium chloride diluted to 100ML Procedure: 1g of clean tissue was taken in a homogenizer contain a few ml of chloroformmethanol reagent and the homogenate was then centrifuged at 3000rpm for 10 minutes. Then contents were transferred into test tubes and a few ml of 0.5N KCl added to the filtrate. The face of KCl was then removed and the lower phase is transferred to the pre-weighed watch glass. Then dried for two days. After the content was weighed and the percentage of lipid present tissue was calculated.

Percentage of lipid=weigh of lipid/weight of tissue\*100

Carbohydrate estimation – Phenol Sulphuric acid method

Standards: sugar of 1 mg/ml stock solution prepared and submitted a carbohydrate standard curve from the following dilution series, used 400,800, 1600, and 2000 $\mu$ l of the stock and made up each sample with dh<sub>2</sub>o to a final volume of 2000 $\mu$ l to make 0.2 $\mu$ g/ $\mu$ l, 0.4ug/ $\mu$ l, 0.6 $\mu$ g/ $\mu$ l 0.8ug/ $\mu$ l, 1ug/ $\mu$ l concentrations, respectively. The blank of (0 $\mu$ g/ $\mu$ l) was prepared as well by just pipetting 2000ul dH<sub>2</sub>o.

To each standard, blank and the sample, added 50  $\mu$ l of 80% (w/v) phenol solution, then vortexed thereby added 2.0 ml concentrated sulphuric acid in a stream then stood for 10 minutes at room temperature. Red absorbance at 490nm, therefore, determined the sugar content of the unknown samples.

Sample	Protein estimation results(mean ± sd)		Carbohydrate estimation results (mean ± sd)			Lipid estimation results Mean ± sd)			
	Fresh fish	Salt dried	P<	Fresh Fish	Salt dried	P<	Fresh	Salt dried	P<
Rastrelliger kanagurta	1.241±0. 895	0.792±0. 088	0.05	0.1395±0 .033	0.084±0.00 85	0.101	1.111±0. 188	0.903	0.0 01
Sardinella longiceps	1.994±0. 086	0.846±0. 687	0.01	0.47±0.0 43	0.38±0.02	0.05	1.74±0.3 41	0.981±0 .213	0.0 01

#### **III. RESULTS AND DISCUSSION Table 1** shows the nutritive analysis of Rastrelliger kanagurta and Sardinella longiceps

The present investigation showed that there is a decrease in the amount of protein in the flesh of salt-dried fish when compared with the flesh of fresh fish. Mahbuba Aktar et.al .,(2011) studied there marine dry fishes (Harpoodon nehereus,johnius dussumieri, and lepturacanthus suvala) and observed that the protein level varied from 58.33%-51.98%,64.39%-56.46%, and 71.90%-67.22% respectively during the storage period. Martin et.al.,(2000)suggested that a decreased quantity of protein is caused by the salt

used for drying.salt affects protein concentration. Lipids are biochemically very important, on account of their role as a chief storage form of energy and also structural molecules of the cell. The present investigation revealed that there is a decrease amount of lipid in the flesh of salt-dried fish when compared with fresh fish.rubbi et al (1987) reported that the lipid content from 0.45%-15.51% in marine fishes is very close to the present investigation. Muhuba, et.al., (2100)observed the mean percentage of lipid in dried fish samples were



varied from 77.78%-6.86%,5.54%-4.87% and 7.79%-6.66% during drying.surono et al (2002) showed that the lipid oxidation in codfish correlated with a decrease in available protein, the greatest reaction occurs in the sample with high salt content. The oxidation of lipids results in alternations in flavor, color, and nutrition. The salt alters the structural integrity of membranes living lipid molecules more accessible to react with ros or with other pro-oxidants fish has an only a small amount of carbohydrates, which insignificant from a nutritional point of view. The present study also shows a decrease in carbohydrate content in salt dried fishes when compared with fresh fish.

#### **IV. CONCLUSION**

The present investigation revealed that there was a decrease in the amount of protein, lipid, and carbohydrate content in salt dried fish when compared to fresh fish. This is because the salt used for drying affects protein, lipid, and carbohydrate present in the tissue. Salt oxidize the protein and lipid. Fish consumption in the fresh condition is more preferable than salt dried fish because of their high nutritive value.

#### REFERENCE

- Adesiyun, A. A. (1993). Prevalence of Listeria spp., Campylobacter spp. Salmonella spp. Yersinia spp. And Toxigenic Escherichia coli on Meat and Seafood in Trinidad. Food Microbiology. 10:395-403.
- [2]. Afolabi, O. A.; Omosola, A. A. And Olusegun, L. O. (1984). Quality Changes of Nigerian Traditionally Processed Freshwater Fish Species. I. Nutritive and Organoleptic Changes. J. Food Technology. 19:333–340.
- [3]. Amirthalingam, C. And M. Y. Khalifa (1965). A. Guide to the common commercial freshwater fishes in Sudan. Government printing press, Khartoum. 197 pp.
- [4]. AOAC (1990). Official methods of the analysis 14th edition. Association of American Analytical chemist, Washington D.C U.S.A.
- [5]. Awouda, F. A. (1988). Studies on the Body composition of Adult Oreochromis niloticus (Trewavas) and Alestes dentex (L.) From White Nile at Khartoum Area with Special Reference to Seasonal Changes and Gonadial Maturity. M. Sc. Thesis, Department of Zoology, University of Khartoum, Sudan 50 pp.
- [6]. Bellagha, S. A. Sahli, A. Farhat, N. Kechaou, and A. Glenza, (2007). Studies on salting and drying of sardine (Sardinella aurita):

Experimental kinetics and modeling. J Food Eng, 78: 947-952.

- [7]. Clucas, I. J. And P. J. Sctcliffe, (1987). An introduction to fish handling and processing. Report of the Tropical Products Institute, pp: 143-186.
- [8]. Clucas, I. J. And Ward, A. R. (1996). Post– Harvest Fisheries Development: A Guide to Handling, Preservation, Processing, and Quality. Natural Resources Institute (NRI), U. K. Connell J. J. (1995). Control of Fish Quality. Fourth edition. Fishing News Book. Austria. Pp 37-64
- [9]. Dambergs, N. (1963). Extractives of Fish Muscle 3. Amounts, Sectional Distribution and Variations of Fat, Water Soluble Protein and Moisture in Cod Gadus morhua (L.) Fillets. J. Fish. Res. Bd. Canada 20 (4):909– 918.
- [10]. FAO (1992a). Fermented Fish in Africa. A Study on Processing, Marketing, and Composition. FAO Fisheries Technical Paper No. 329, Rome, Italy. 57 pp.
- [11]. FAO (1992b). Manual of Food Quality Control 4 Rev. 1: Microbiological Analysis. Food and Agriculture Organization of the United Nations, Rome, Italy.
- [12]. Horner, W. F. A. (1997). Salting. In: Fish Processing Technology: Preservation of Fish, Curing. GM Hall (ed), Chapman & Hall Publishers. UK, pp: 32-72. Huss, H. H. (1988). Fresh Fish Quality and Quality Changes. FAO Fisheries Series No. 29 Rome, Italy. 132 pp.
- [13]. Jackson, R. I. (1971). The Importance of Fish Inspection in the Rational Utilization of Fishery Resources. In Kreuzer, R. (ed.) "Fish Inspection and Quality Control" Fishing News (Books) Limited, London pp. 2–6.
- [14]. Jay, J. M. (1992). Modern Food Microbiology. Microbiological Indicators of Food Safety and Quality, Principles and Quality Control, and Microbiological Criteria. New York: Van Nostrand Reinhold.
- [15]. Lunven, P. (1982). The Role of Fish in Human Nutrition. FAO Food and Nutrition, 8 (2):9–18.
- [16]. Mahmoud, Z. N. (1977). Studies on Meat Quality of some Nile Fish. M. Sc. Thesis, Department of Zoology, University of Khartoum, Sudan, 65 pp.
- [17]. Omer, A. S. (1984). Preliminary Studies on the Chemical Composition of the Flesh of Hydrocyon forskali (Cuvier and Valenciennes, 1868) B. Sc. Honors



dissertation Department of Zoology, University of Khartoum, Sudan, 38 pp.

- [18]. Pearson, D. (1976). The Chemical Analysis of Foods. 7th edition, Churchill, Livingstone, Edinburgh, London, and New York .
- [19]. Saito, H., and Udagawa, M. (1992). Assessment of Oxidative Deterioration of Salted Dried Fish by Nuclear Magnetic Resonance, JAOCS, Vol. 69 (11):1157– 1159.
- [20]. Wheaton, F., and Lawson, T. (1985). Processing Aquatic Food Products. New York. 517 pp.
- [21]. Wilson (2007). Nutrition and feeding of fish aquaculture, 267: 1–2.

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